

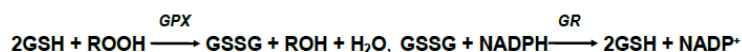
Glutathione Peroxidase Assay Kit (Colorimetric)

LS-K197-100 (100 Tests) • Store at -20°C



Introduction

GLUTATHIONE PEROXIDASE represents an enzyme family with peroxidase activity whose main biological role is to protect the organism from oxidative damage. It helps prevent lipid peroxidation of cellular membranes by removing free peroxide in the cell. GPX catalyzes the following reaction with glutathione reductase



Simple, direct and high-throughput assays for GPX activity find wide applications. LSBio's improved assay directly measures NADPH consumption in the enzyme coupled reactions. The measured decrease in optical density at 340 nm is directly proportional to the enzyme activity in the sample.

Key Features

- Sensitive and accurate. Use 10 µL sample. Linear detection range 40 to 800 U/L GPX activity.

Applications

- Direct Assays: GPX activity in biological samples.
- Drug Discovery/Pharmacology: effects of drugs on GPX activity.

Components

Component	K197-100
	100 Tests
Assay Buffer	25 mL
GR Enzyme	250 µL
Glutathione	Dried
NADPH	Dried
Calibrator	Dried
Peroxide Solution	50 µL

Materials Not Supplied

Pipetting devices, centrifuge tubes, clear flat-bottom uncoated 96-well plates, plate reader capable of reading optical density at 340 nm every minute, homogenizer, etc.

Storage

The kit is shipped on ice. Store all kit components at -20 °C.

FOR RESEARCH USE ONLY! Not for use in humans.

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www.LSBio.com • (206) 464-1554 • TechnicalSupport@LSBio.com

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Sample Preparation

All samples should be clear and free of any turbidity or particles. Liquid samples (e.g. non-hemolyzed serum, plasma) can be assayed directly.

Homogenize tissue (10 mg) and cells (10⁶) in 200 μ L cold PBS and then centrifuge 10 min at 14,000 rpm to pellet any debris. Use the clear supernatant for the assay. If not assayed immediately, freeze supernatant at -80°C (stable for 1 month).

Assay Procedure

1. Reagent Preparation. Equilibrate all components to room temperature. Briefly centrifuge all tubes before opening. Reconstitute the NADPH tube with 400 μ L dH₂O (final 35 mM).
Reconstitute the Glutathione tube with 540 μ L dH₂O (final 100 mM). Reconstitute the Calibrator with 100 μ L dH₂O (final 100 mM). Vortex tubes to mix. Keep these reconstituted reagent tubes on ice. Unused reagents are stable for three weeks when stored frozen at -20°C.
2. Standards and Samples. Mix 12 μ L of the Calibrator with 188 μ L dH₂O (equivalent to 6 mM NADPH). Dilute this calibrator stock as shown in the Table below.

No	6 mM Calibrator + H ₂ O	Vol (μ L)	[Equiv. NADPH] (mM)
1	100 μ L + 0 μ L	100	6.0
2	60 μ L + 40 μ L	100	3.6
3	30 μ L + 70 μ L	100	1.8
4	0 μ L + 100 μ L	100	0

Transfer 10 μ L standards into wells of a clear flat-bottom 96-well plate. Add 190 μ L Assay Buffer to all standard wells. Transfer 10 μ L of each sample into separate wells of the 96-well plate. In addition, for each assay run, include a background control that only contains 10 μ L Assay Buffer.

Note: For unknown samples, perform several dilutions to ensure that GPX activity is within the linear range of 40 to 800 U/L.

3. Assay. Prepare enough Working Reagent for all Sample and Control wells by mixing, for each well, 90 μ L Assay Buffer, 5 μ L Glutathione, 3 μ L 35 mM NADPH and 2 μ L GR enzyme. Add 90 μ L Working Reagent quickly to the Sample/Control wells. Tap plate to mix.

Transfer 5 μ L Peroxide Solution into an EMPTY 1.5-mL Eppendorf tube. Add 1495 μ L dH₂O and mix well by vortexing for at least 30 seconds. Immediately dilute this solution 1:10 in dH₂O to generate the 1X Substrate Solution. Use the diluted Solutions within one hour.

With a multi-channel pipette, add 100 μ L 1X Substrate Solution to all Sample and Control wells. Tap plate quickly to mix well contents thoroughly. Immediately read OD_{340nm} (time zero, OD₀) and again at 4 min (OD₄).

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Calculations

Use OD values at 4 min for NADPH standards. Subtract blank value (#4) from the standard values. Plot the ΔOD against standard concentrations and determine the slope of the standard curve. Calculate the $\Delta OD_s = (OD_0 - OD_4)$ for the samples and $\Delta OD_B = (OD_0 - OD_4)$ for the background control. Calculate the GPX activity of Sample,

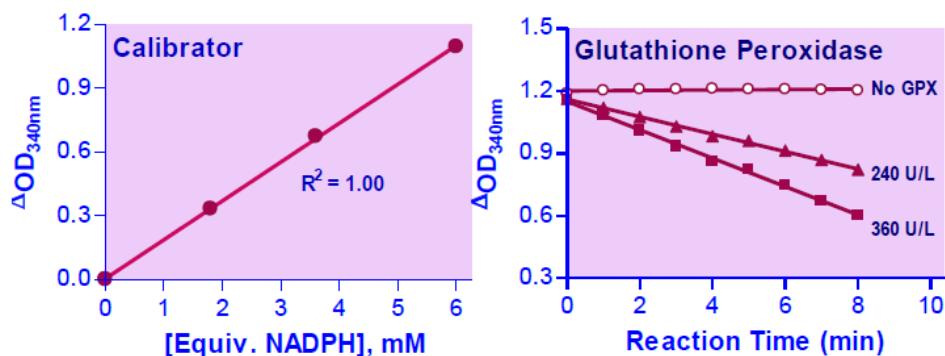
$$\text{GPX Activity (U/L)} = \frac{\Delta OD_s - \Delta OD_B}{\text{Slope (mM}^{-1}) \times 4 \text{ (min)}} \times 1000 \times n$$

The factor 1000 converts mmoles to μ moles. n is the sample dilution factor.

Note: if calculated GPX activity is higher than 800 U/L, or initial OD_{340nm} is >1.5 in sample wells, dilute sample in dH₂O and repeat assay. Multiply the results by the dilution factor.

Unit definition: one unit is the amount of GPX that produces 1 μ mole of GS-SG per min at pH 7.6 and room temperature.

Sample Data



Version: V.08.09.2018

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