

D-Lactate Assay Kit (Fluorometric)

LS-K237-100 (100 Tests) • Store at -20°C



Introduction

Lactate is generated by lactate dehydrogenase (LDH) under hypoxic or anaerobic conditions. Monitoring lactate levels is, therefore, a good indicator of the balance between tissue oxygen demand and utilization and is useful when studying cellular and animal physiology. D-Lactate is produced in only minor quantities in animals and measuring for D-lactate in animal samples is a means to determine the presence of bacterial infection. Simple, direct and automation-ready procedures for measuring lactate concentration are very desirable. This lactate assay kit is based on lactate dehydrogenase catalyzed oxidation of lactate, in which the formed NADH reduces a probe into a highly fluorescent product. The fluorescence intensity of this product, measured at $\lambda_{ex/em} = 530/585$ nm, is proportional to the lactate concentration in the sample.

Key Features

- Sensitive and accurate. Detection limit of 1 μ M and linearity up to 50 μ M D-lactate in 96-well plate assay.
- Convenient. The procedure involves adding a single working reagent, and reading the fluorescence after 60 min. Room temperature assay.
- High-throughput. Can be readily automated as a high-throughput 96-well plate assay for thousands of samples per day.

Applications

- Direct Assays: D-lactate in serum, plasma, urine, cell media samples and other biological samples.

Components

Component	K237-100
	100 Tests
Assay Buffer	10 mL
Enzyme A	120 μ L
Enzyme B	120 μ L
NAD Solution	1 mL
Probe	750 μ L
Standard	1 mL

Materials Not Supplied

Pipetting (multi-channel) devices. Black, flat bottom 96-well plates and fluorescent plate reader capable of reading at $\lambda_{ex/em} = 530/585$ nm.

Storage

The kit is shipped on ice. Store all reagents at -20°C. Shelf life: 6 months after receipt.

FOR RESEARCH USE ONLY! Not for use in humans.

LifeSpan BioSciences, Inc. • 2401 Fourth Avenue, Suite 900, Seattle, WA 98121

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Assay Procedure

Sample Preparation

The following substances interfere and should be avoided in sample preparation: EDTA (>0.5 mM), ascorbic acid, SDS (>0.2%), sodium azide, NP-40 (>1%) and Tween-20 (>1%). Samples suspected of having endogenous L-LDH enzyme activity (e.g. serum, plasma, culture medium with FBS, etc.) should be deproteinated using a 10 kDa spin filter (e.g. Microcon YM-10). Deproteinated serum should be diluted 3 × with dH₂O. Samples containing higher than 50 μM pyruvate require an internal standard.

Procedure

1. Standard Curve. Prepare 1000 μL 40 μM D-lactate Premix by mixing 2 μL 20 mM Standard and 998 μL distilled water. For cell culture samples, prepare 1000 μL 40 μM D-lactate Premix by mixing 2 μL 20 mM Standard and 998 μL culture medium without serum. Dilute standard as follows.

No	Premix + H ₂ O or Medium	D-Lactate (μM)
1	100 μL + 0 μL	40
2	60 μL + 40 μL	24
3	30 μL + 70 μL	12
4	0 μL + 100 μL	0

Transfer 50 μL standards into wells of a black, flat bottom 96-well plate.

Samples. Add 50 μL sample to two separate wells. Set up two reactions for each sample: one with added Enzyme A (Sample) and a No Enzyme A control (Sample Blank).

Samples requiring an internal standard, will need three separate reactions: 1) Sample plus Standard, 2) Sample alone and 3) Sample Blank. For the internal standard first prepare 400 μL 250 μM D-lactate standard by mixing 5 μL 20 mM Standard and 395 μL dH₂O. For the Sample plus Standard well, add 5 μL 250 μM D-lactate and 45 μL sample. For the Sample and Sample Blank wells, add 5 μL dH₂O and 45 μL sample.

2. Reagent Preparation. Spin the Enzyme tubes briefly before pipetting. For each Sample and Standard well, prepare Working Reagent by mixing 40 μL Assay Buffer, 1 μL Enzyme A, 1 μL Enzyme B, 10 μL NAD and 5 μL Probe. Fresh reconstitution is recommended. For the Sample Blanks, the Working Reagent includes 40 μL Assay Buffer, 1 μL Enzyme B, 10 μL NAD and 5 μL Probe (NO Enzyme A).
3. Reaction. Add 50 μL Working Reagent per reaction well quickly. Tap plate to mix briefly and thoroughly. Incubate for 60 min at RT protected from light.
4. Read fluorescence $\lambda_{\text{ex/em}} = 530/585 \text{ nm}$.

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Calculations

Plot the D-lactate Standard Curve and determine its slope. The D-lactate concentration of the sample is computed as follows:

$$[\text{D-Lactate}] = \frac{F_{\text{SAMPLE}} - F_{\text{BLANK}}}{\text{Slope } (\mu\text{M}^{-1})} \times n \quad (\mu\text{M})$$

where F_{SAMPLE} and F_{BLANK} are the fluorescence intensity values of the Sample and Sample Blank respectively. Slope is the slope of the standard curve and n is the dilution factor (e.g. $n = 3$ for serum samples).

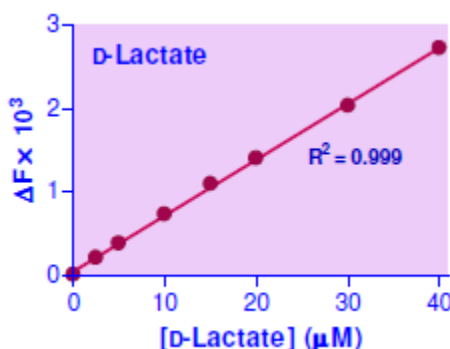
If an internal standard was needed, the sample D-lactate concentration is computed as follows:

$$[\text{D-Lactate}] = \frac{F_{\text{SAMPLE}} - F_{\text{BLANK}}}{F_{\text{STANDARD}} - F_{\text{SAMPLE}}} \times 27.8 \quad (\mu\text{M})$$

where F_{SAMPLE} and F_{BLANK} are the fluorescence intensity values of the Sample and Sample Blank respectively and F_{STANDARD} is the fluorescence intensity value of the Sample plus Standard.

Note: if the sample ΔF value is higher than the ΔF for 40 μM D-lactate standard or greater than the ΔF for the internal standard, dilute the sample in water and repeat the assay. Multiply the results by the dilution factor.

Sample Data



Standard Curve in 96-well plate assay in water.

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